

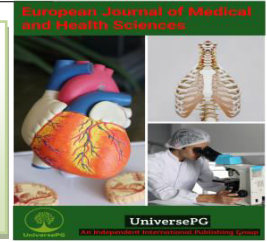


Publisher homepage: www.universepg.com, ISSN: 2663-7529 (Online) & 2663-7510 (Print)

<https://doi.org/10.34104/ejmhs.020.01010106>

European Journal of Medical and Health Sciences

Journal homepage: www.universepg.com/journal/ejmhs



Microbiological Screening and Antimicrobial Sensitivity Profiling of Wound Infections in a Tertiary Care Hospital of Bangladesh

Abdullah Akhtar Ahmed^{1&2}, Nusrat Akhtar Juyee³, S.M. Ali. Hasan⁴, and Mohammad Zakerin Abedin^{1*}

¹Dept. of Microbiology, Khwaja Yunus Ali University, Sirajganj, Bangladesh; ²Dept. of Microbiology, Khwaja Yunus Ali Medical College, Sirajganj, Bangladesh; ³Dept. of Microbiology and Immunology, Bangabandhu Sheikh Mujib Medical University, Dhaka, Bangladesh; and ⁴Dept. of Gastroenterology, Bangabandhu Sheikh Mujib Medical University, Dhaka, Bangladesh.

*Correspondence: zakerin.du2016@gmail.com (Mohammad Zakerin Abedin, Assistant Professor and Head, Dept. of Microbiology, Khwaja Yunus Ali University, Sirajganj, Bangladesh).

ABSTRACT

Wound infection is a major problem in hospitals in developing countries. Wound infection causes morbidity and prolonged hospital stay thus this prospective study was conducted for a period of seven months (January 2019 to July 2019). A total of 217 specimens (wound swabs and pus exudates) from wound infected patients in a Tertiary Care Hospital in Bangladesh. A retrospective study of the microbiological evaluation was done by cultural growth as well as Gram staining and biochemical examination to identify the bacterial isolates. Finally, the antimicrobial vulnerability testing was performed by Kirby-Bauer disc diffusion conventional method. A total of 295 samples were tested. Out of which 217 (73.5%) were found culture positive. *E. coli* was the most predominant gram-negative isolates whereas *Staphylococcus aureus* and Coagulase-negative *Staphylococcus* were the most commonly isolated gram-positive organisms. Antimicrobial sensitivity profile of bacterial isolates revealed imipenem, meropenem, amikacin, and nitrofurantoin to be the most effective antimicrobials against gram-negative isolates, whereas imipenem, meropenem, amikacin, nitrofurantoin, amoxiclav, and gentamicin were the most effective drugs against gram-positive isolates. The result of this examination contributes to the identification of basic causative microbes involved in wound infection and findings of antibiotic susceptibility patterns can be helpful for primary care physicians to optimize the treatment modalities, articulate policies for empiric antimicrobial therapy, and to minimize the rate of infection among wound infected patients.

Keywords: *Staphylococcus aureus*, Screening, *Pseudomonas aeruginosa*, and Antimicrobial sensitivity profiling.

INTRODUCTION:

The wound infection is known as the presence and growth of microbes in wound (Howell-Jones *et al.*, 2005). The development of wound infection depends on the integrity and prospective function of skin (Sorg *et al.*, 2017). The local wound conditions, systemic host defenses, microbial burden, and existing ailments just as outward factors, for example, pre, intra and

postoperative care are the potential variables for infection of a patient. Along these lines, it is hard to foresee which wound will get infected. The general frequency of wound sepsis is from 10-33% (Neelima *et al.*, 2013). Relative protection from anti-microbials nearly more harmful strains and ability to adjust rapidly to changing climate make the microorganisms obtained in medical clinics a matter of concern

(Plummer *et al.*, 2004). Notwithstanding current careful methods and the utilization of anti-infection prophylaxis, Surgical Site Infection (SSI) is one of the most widely recognized intricacies experienced in medical procedure. SSI places a critical significant on both the patient and wellbeing framework. SSI is thus a major cause of morbidity, prolonged hospital stay and increased health costs (Mezemir *et al.*, 2020)

Wound infection therapy with antibiotics and ideal treatment regimens remains under documented. There are many published guidelines are for the most part dependent on expert conclusion instead of proof based information. The choice of appropriate antimicrobial drugs has been uncertain. Despite the fact that prophylactic utilization of antimicrobials can help decrease the danger of disease and advances wound mending, it's anything but an immediate substitute for good nearby twisted consideration, for example, water system and careful debridement. In addition, supported utilization of antimicrobials decreases the improvement of antimicrobial obstruction (AMR) (Elbur *et al.*, 2013; Rijal *et al.*, 2017).

It has been revealed by reviewing of recent practices that potentially inappropriate and inconsistent use of antimicrobials following surgical procedures contributes to development of AMR. Likewise, fittingness of the circumstance, the span, course, and determination of these agents stays elusive (Tweed *et al.*, 2005 and Landis *et al.*, 2008). In Bangladesh, the efficient review and Meta analysis is planned to give cross country pooled estimates of microbial profiles, wound culture positivity, and AMR patterns of wound disease. This will manage as a benchmark for creating antimicrobial reconnaissance programs and producing proof based determination of antimicrobials to protect the accessible antimicrobials and contain AMR.

MATERIALS AND METHODS:

Study Place - This study was conducted in Microbiology section of the Department of Laboratory Services, Khwaja Yunus Ali Medical College Hospital of remote region Enayetpur of Bangladesh from January 2019 to July 2019.

Sample Collection and Processing - All the wounds were judged as infected by the presence of purulent

material. Before wound cleansing and dressing conducted pus discharge from 92 and wound swab from 203 patients were collected. Two wound swabs were collected from the wound and from a drop of aspirate using sterile cotton tipped applicators. First swab used to make smear on a clean glass slides and Gram staining was done for direct microscopic examination at 100X objective to know details bacteria-l morphology and leucocytes cells.

Isolation and Identification - The second swab or drop of aspirate pus was utilized for culture by sterile inoculating loop on routine laboratory culture media like Nutrient Agar, Blood Agar, and MacConkey's agar, and then incubated at 37° C for 24 hrs to 48 hours aerobically.

Isolates were confirmed on standard microbiological methods. They included morphological and cultural characteristics, Gram staining, motility, hydrogen sulphide production, indole production, carbohydrate fermentation test, and Simon citrate utilization test. Other tests include catalase test, oxidase test, coagulase, haemolysis on blood agar, morphological and cultural characteristics on eosin-methylene blue agar, and mannitol salt agar.

Antibiotic Sensitivity - The antibiotic sensitivity pattern was done by Potentiated Disc Diffusion Test (PDT) using Muller Hinton agar through following Kirby Bauer's method as per standard CLSI guidelines (CLSI 2017). Commonly used antibiotics (Oxoid antibiotic disc, UK) were tested against both of both Gram positive and Gram negative bacterial isolates.

Exclusion Criteria - Patients already on antibiotics were excluded from the study

Data Analysis - The SPSS version 16.0 was used for analysis of data. The percentage of frequencies were generated for different categorical variables such as rate of isolation, type of bacteria, rate of antibiotic sensitivity, resistance, intermediate of the organisms.

RESULTS:

A total of 295 samples received in the laboratory were randomly selected for this study. Out of which 217 samples showed aerobic growth, and 78 remained sterile even after 48 hours incubation at 37°C. Among

217 growth positive cases, 203 (68.8%) samples were from pus swab and 92 (31.2%) were from aspirate. Among 78 negative cases, 49 (62.8%) samples were from pus swab and 29 (37.2%) were from aspirate (Table 1 and Fig 1).

Table 1: Frequency of growth and no growth patterns in total wound samples.

Types of sample	Growth		No Growth		Total number (%)
	Number (N)	Percent	Number (N)	Percent	
Pus swab	154	75.9 %	49	24.1%	203 (68.8%)
Pus aspirate	63	68.5 %	29	31.5%	92 (31.2%)
Total positive cases	217		Total negative cases	78	295 (100%)

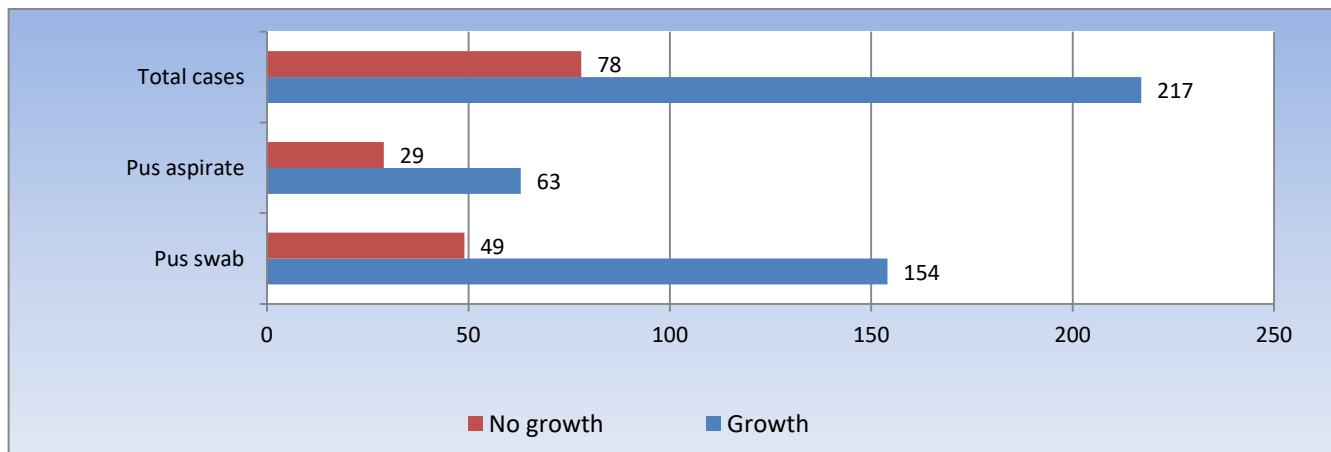


Fig 1: Total number of frequency of wound infection of the suspected patients.

Table 2: Distribution Pattern of Bacterial Isolates of wound infection (n= 217).

Bacterial isolates		Total No.	Percentage
Gram negative bacteria n=121 (55.8%)	<i>Escherichia coli</i>	89	41
	<i>Pseudomonas aureginosa</i>	26	12
	<i>Klebsiella spp</i>	06	3
Gram positive bacteria n=96 (44.2%)	<i>Staphylococcus aureus</i>	68	31
	Coagulase Negative <i>Staphylococcus</i>	28	13
Total		217	100

Majority of the wound was infected with single organism. A Gram negative *Bacillus* was 55.8% and Gram positive *Cocci* 44.2%. Most frequently isolated organisms were *E. coli*, 89(41%) followed by *Staphylococcus aureus* 68(31%), Coagulase negative *Staphylococcus*, other isolates included *Klebsiella spp* 6(3%) and *Pseudomonas aureginosa* 26(12%), (Table 2).

High number of samples (45) were collected from patients of age group 41 - 50 years and the least i.e. 09 samples were obtained from patients of age group 0 - 10 years. Among 217 positive cases, the highest posi-

tive cases 118 (54.4%) were male and rest 99 (45.6%) were female (Table 3).

Antibiotic susceptibility pattern of the isolates showed imipenem 202 (93.1%), meropenem 175 (80.6%) and amikacin 166 (76.5%) were found effective for both Gram-negative and Gram-positive isolates (Shahen et al., 2019). Cephadrine 48 (22.1%) was found to be weak in inhibiting both Gram-positive and gram-negative bacterial growth. For Gram-negative isolates Cephadrine showed highest resistant pattern (Table 4).

Table 3: Gender and age-wise distribution of wound infected patients (n=217).

Age group (in years)	Male		Female		Total (no. of cases)	Percentage % (n=217)
	No. of cases	% age	No. of cases	% age		
0-10	5	4.2	4	4.1	9	4.0
11-20	8	6.8	11	11.1	19	8.7
21-30	23	19.5	23	23.2	46	21.1
31-40	19	16.1	22	22.2	41	19.0
41-50	31	26.3	14	14.1	45	21.0
51-60	22	19.4	16	16.2	38	17.5
>60	10	8.5	9	9.1	19	8.7
Total	118	54.4	99	45.6	217	100

Table 4: Antimicrobial sensitivity profile of bacterial isolates from the wound patients.

Antibiotics	Bacterial isolates, Number (n) with percentage (%)					
	<i>E.coli</i> 89 (41)	<i>S. aureus</i> 68 (31)	NoSA 28 (13)	<i>P. aureginosa</i> 26 (12)	<i>Klebsiella spp</i> 06 (3)	Total 217(100)
Amoxiclav	22 (24.7)	40 (58.8)	20 (71.4)	04 (15.3)	04 (66.6)	90 (41.5)
Amikacin	60 (67.4)	58 (85.2)	24 (85.7)	19 (73)	05 (83.3)	166 (76.5)
Azithromycin	18 (20.2)	13 (19.1)	07 (25)	15 (57.6)	03 (83.3)	56 (25.8)
Ceftazidime	21 (23.5)	17 (25)	17 (60.7)	19 (73)	04 (66.6)	78 (35.9)
Ceftriaxone	21 (23.5)	30 (44.1)	20 (71.4)	18 (69.2)	03 (83.3)	92 (42.4)
Cefuroxin	15 (16.8)	28 (41.1)	20 (71.4)	02 (7.7)	02 (33.3)	67 (30.8)
Cephadrine	05 (5.6)	23 (33.8)	17 (60.7)	02 (7.7)	01 (16.6)	48 (22.1)
Ciprofloxacin	17 (19.1)	25 (36.7)	13 (46.4)	16 (61.5)	04 (66.6)	75 (34.5)
Gentamicin	36 (40.40)	40 (58.8)	27 (96.4)	14 (53.8)	05 (83.3)	122 (56.2)
Imipenem	79 (88.7)	65 (95.5)	28 (100)	23 (88.4)	06 (100)	202 (93.1)
Meropenem	68 (76.4)	56 (82.3)	23 (82.1)	23 (88.4)	05 (83.3)	175 (80.6)
Levofloxacin	18 (20.2)	31(45.5)	13 (46.4)	16 (61.5)	05 (83.3)	83 (38.2)
Nitrofurantoin	48 (53.9)	46 (67.6)	23 (82.1)	04 (15.3)	02 (33.3)	123 (56.7)

DISCUSSION:

Wound contamination has been a significant worry among medical services specialists not just regarding expanded injury to the patient yet additionally taking into account its weight on money related assets and the expanding necessity for savvy the board inside the medical care framework. To take care of the serious issue of medical care framework this investigation is completed to discover the bacteriological profile of

wound contaminations and antimicrobial power lessness example of secludes. It was seen that the commonest age bunch influenced is 21-30 years which are connected with the examinations done by Malik *et al.*, 2011 and Afroz *et al.*, 2015. Regarding the sex distribution of the patients in the present study, males (54.4%) were affected more than females (45.6%). The possible reasons for this male preponderance can be related to socio-economic and cultural habits of

earning the livelihood primarily by males and also to their adventurous nature and the greater desire to be active in comparison to their female counterparts (Kai-Yang *et al.*, 2008). This study was correlated with Ramesh *et al.* (2013) which showed males (60%) more affected than females, Sowmya *et al.* (2014) (66.6%) and Malik *et al.* (2011) (51.9%) also showed the predominance of males over females.

In this study on wound infection five different types of organisms were identified. Highest percentage of isolation was *Escherichia coli* (41%), followed by *Staphylococcus aureus* (31%), Coagulase Negative *Staphylococcus* (13%), *Pseudomonas aeruginosa* (12%) and *Klebsiella pneumoniae* (3%). Similarly, *Escherichia coli* (55.9%) in highest percentage were detected in Dhaka, Bangladesh study on wound infection where four different types of organisms were identified (Atiyeh *et al.*, 2007). However, *Staphylococcus aureus* (27.5%) was found as the most common isolate followed by CoNS (8.5%), (Tapan *et al.*, 2013). The antibiogram studies indicate the emergence of extensively drug resistant and pandrug resistant strains. The isolates were exhibited resistance to the commonly used antibiotics as well as new generation antibiotics. In our study, most of the isolated pathogens were found sensitive to costly antibiotics like Imipenem (93.1%), followed by Meropenem (80.6%), Amikacin (76.5%) and least effective cheap antibiotics were Ampicillin, Amoxi-cillin, Cefixim and Cefotaxime. Amikacin was also found highly sensitive to wound infection pathogens by Neelima *et al.* 2013. In contrast to our study, the incidence of resistant to Meropenem (83.78%), Imipenem (85.41%) and Amikacin (87.03%) were found by Abedin *et al.* (2020).

CONCLUSION:

This experiment was completed to decide bacteriological and antimicrobial affectability profile of wound contaminations in a tertiary care hospital of Bangladesh. This study may update the physicians in the different antimicrobial choices accessible in the treatment of wound diseases, consequently assisting with diminishing morbidity and mortality in long haul; it might decrease the expense of treatment. In our investigation assortment of aerobic bacteria are

isolated from wound with prevalence of *Escherichia coli* followed by *Staphylococcus aureus*. Imipenem was the most delicate medication among both Gram positive and Gram negative bacteria and later on Amikacin and Ceftazidime was least effective (Abedin *et al.*, 2020). It tends to be concluded that more complete investigations are needed every once in a while to characterize the extent of issue and produce information for strategy choice on ideal medication modalities.

ACKNOWLEDGEMENT:

We acknowledge the staff of the Department of Laboratory Services, Khwaja Yunus Ali Medical College Hospital and Department of Microbiology, School of Biomedical Science, Khwaja Yunus Ali University, Sirajganj, Bangladesh.

CONFLICTS OF INTEREST:

Authors declare that no competing interest exists to publish the present research work.

REFERENCE:

1. Abedin MZ *et al.* (2020). Enumeration of the antimicrobial susceptibility patterns of different bacterial isolates from ENT patients with ear infections, *Eur. J. Med. Health Sci.*, 2(4), 68-73.
<https://doi.org/10.34104/ejmhs.020.068073>
2. Abedin MZ, Aktar MB, Zaman MSU, Jarin L, Miah MAS, and Ahmed AA, *et al.* (2020). Predominance of nosocomial pathogens among patients with post-operative wound infections and evaluation of the anti-biotic susceptibility patterns in rural hospitals in Bangladesh. *Recent Adv Biol Med.*; 6(4): 9800005.
<https://doi.org/10.18639/RABM.2020.9800005>
3. Afroz Z, Metri BC, Jyoti P. (2015). Bacteriological Profile and Antimicrobial Susceptibility Pattern of Skin and Soft Tissue Infections among Gram Negative Bacilli in a Tertiary Care Hospital of South India. *J. Pharm. Scie & Res.* 7(7): 397-400.
<https://www.jpsr.pharmainfo.in/Documents/Volumes/vol7Issue07/jpsr07071502.pdf>

4. Atiyeh BS *et al.* (2007). Effect of silver on burn wound infection control and healing: review of the literature. *Burns*. **33**(2): 139-48. <https://doi.org/10.1016/j.burns.2006.06.010>
5. CLSI, (2017). Performance Standards for antimicrobial susceptibility testing, twenty-third information supplement. *M100-S24*. **34**: 50-57.
6. Elbur AI, Yousif M, Abdel-Rahman ME. (2013). Prophylactic antibiotics and wound infection. *J Clin Diagn Res*. **7**(12): 2747.
7. Howell-Jones RS *et al.* (2005). A review of the microbiology, antibiotic usage and resistance in chronic skin wounds, *Journal of Antimicrobial Chemotherapy*. **55**(2): 143-149. <https://doi.org/10.1093/jac/dkh513>
8. Landis SJ. (2008). Chronic wound infection and antimicrobial use. *Adv Skin Wound Care*. **21**(11): 531-40.
9. Malik S, Gupta A, Singh KP, Agarwal J, Singh M. (2011). Antibiogram of Aerobic Bacterial Isolates from Post Operative wound Infections at a Tertiary Care Hospital in India. *J. Infect. Dis-Antimicrob Agents*. **28**; 45-51.
10. Mezemir, R., Seid, A., Gishu, T., Demas, T., & Gize, A. (2020). Prevalence and root causes of surgical site infections at an academic trauma and burn center in Ethiopia: a cross-sectional study. *Patient safety in surgery*, **14**, 3. <https://doi.org/10.1186/s13037-019-0229-x>
11. Neelima *et al.* (2013). Bacteriological profile of wound infection in rural hospital in R.R District. *Int J Med Res Heal Sci*. **2**(3):469-473.
12. Olson M *et al.* (1984). Surgical wound infections of 5 year prospective study of 20, 193 wounds at Minneapolis Va Medical Centre. *Ann Surg*. **199**(3): 253-259.
13. Plummer D. (2004). Surgical Wound infections as a performance indicator: agreement of common definitions of wound infections in 4773 patients. *BMJ*. **329**: 720-22. <https://doi.org/10.1136/bmj.38232.646227.DE>
14. Rao R *et al.*, (2013). Bacteriology of Post-Operative wound infections. *Int. J. Pharm Biomed Res*. **4**(2), 72-76.
15. Rijal BP, Satyal D, Parajuli NP. (2017). High burden of antimicrobial resistance among Bacteria causing pyogenic wound infections at a tertiary Care Hospital in Kathmandu, Nepal. *J Pathogens*. **2017**: 7.
16. Shahen MZ, Mahmud S, Uddin ME and Alam MS. (2019). Effect of antibiotic susceptibility and inhibitory activity for the control of growth and survival of microorganisms of extracts of *Calendula officinalis*, *Eur. J. Med. Health Sci*. **1**(1), 1-9. <https://doi.org/10.34104/ejmhs.0190109>
17. Sorg H, Tilkorn D, J, Hager S, Hauser J, Mirastschijski U. (2017). Skin Wound Healing: An Update on the Current Knowledge and Concepts. *Eur Surg Res*. **58**: 81-94. <https://doi.org/10.1159/000454919>
18. Sowmya N *et al.* (2014). A two-year study of spectrum of bacterial isolates from wound infections by aerobic culture and their antibiotic pattern in a tertiary care centre. *Int. J.Cure. Microbiol, App. Sci*. **3**(8): 292-295.
19. Tapan Kr Mandal, Rajeshwari Surpur, Achut Rao, (2013). Dept. Of Microbiology, Navoday Medical College, Raichur: 36th National conference souvenir. Pp. 108.
20. Tweed C. (2005). Prevention of surgical wound infection: prophylactic antibiotics in colorectal surgery. *J Wound Care*. **14**(5): 202-5.
21. Yang KL *et al.* (2008). Epidemiology of pediatric burns requiring hospitalization in China: a literature review of retrospective studies. *Pediatrics*, **122**: 132-42. <https://doi.org/10.1542/peds.2007-1567>

Citation: Ahmed AA, Juyee NA, Hasan SMA, and Abedin MZ. (2020). Microbiological screening and antimicrobial sensitivity profiling of wound infections in a tertiary care hospital of Bangladesh, *Eur. J. Med. Health Sci.*, 2(5), 101-106. <https://doi.org/10.34104/ejmhs.020.01010106> 